RayBio® Glycan Array 100

Patent Pending Technology
User Manual (revised June 24, 2015)

Identification of the specific glycan binding proteins in serum, plasma, cell culture supernatants, cell/tissue lysates or other body fluids.

Cat# GA-Glycan-100-1 (4 Sample Kit)
Cat# GA-Glycan-100-2 (8 Sample Kit)
Cat# GA-Glycan-100-4 (16 Sample Kit)

Please read this manual carefully
before starting your experiment

RayBiotech
The protein array pioneer

ISO 13485 Certified

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TABLE OF CONTENTS

I. Overview 1
Introduction 2
How It Works 3
II. Materials Provided 4
III. General Considerations 5
A. Label-Based vs. Sandwich-Based Method 5
B. Preparation of Samples 5
C. Handling Glass Slides 6
D. Incubation 7
IV. Protocol 8
A. Dialysis of Sample 8
B. Biotin-labeling Sample 9
C. Dry the Glass Slide 10
D. Blocking and Incubation 10
E. Incubation with Detection Antibody Cocktail 12
F. Incubation with Cy3 Equivalent Dye-Streptavidin 12
G. Fluorescence Detection 13
H. Data Analysis 15
V. Glycan Array Map 17
VI. Interpretation of Results and Two Examples 19
VII. Troubleshooting Guide 23

I. Introduction

Glycocalyx, literally meaning 'sugar coat', is an extracellular polymeric coating surrounding many prokaryotic and eukaryotic cells consisting of glycoproteins, glycolipids, proteoglycans and glycosaminoglycans. The constituents of the glycocalyx play an important role in the process of cell signaling, virus transfection, and immunity. However, detection tools for the research of glycobiology are currently in very limited supply.

Raybiotech has pioneered the development of antibody arrays which are now widely applied in the research community with thousands of peer reviewed publications, including Cell and Nature. Taking advantage of advancements in microarray technology developed for antibody arrays, we have developed the largest commercially available glycan array for screening protein-carbohydrate interactions. This array will help researchers: 1) identify the glycans binding partners in biological samples, 2) identify whether target proteins are carbohydrate binding proteins, 3) probe binding of viruses and whole cells to glycans,[1-3] 4) profile the substrate specificity of enzymes (glycosyltransferases, glycosidases, etc.);[4-6] 5) profile the inflammatory immune response.

The 100 synthetic glycans featured in the Glycan Array 100 are the most frequently identified structures showing important binding function in the literature. For example, influenza virus binds to a variety of sialosides with a serotype-specific pattern.[7-9] Galectins, which are involved in apoptosis, cell adhesion and T-cell activation suppression, function by binding beta-galactosides.[10-12]

References

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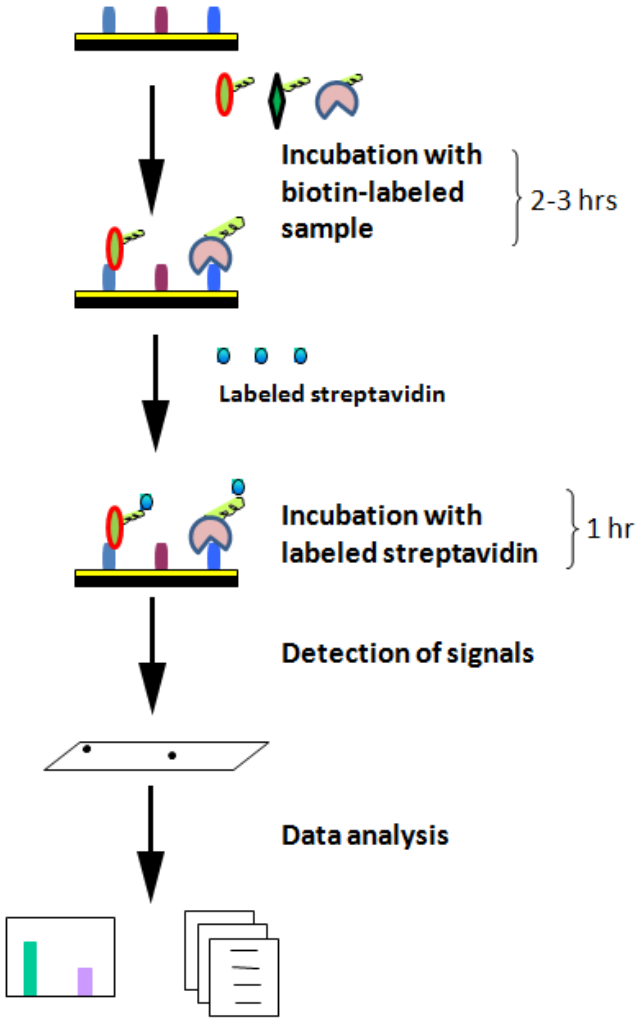
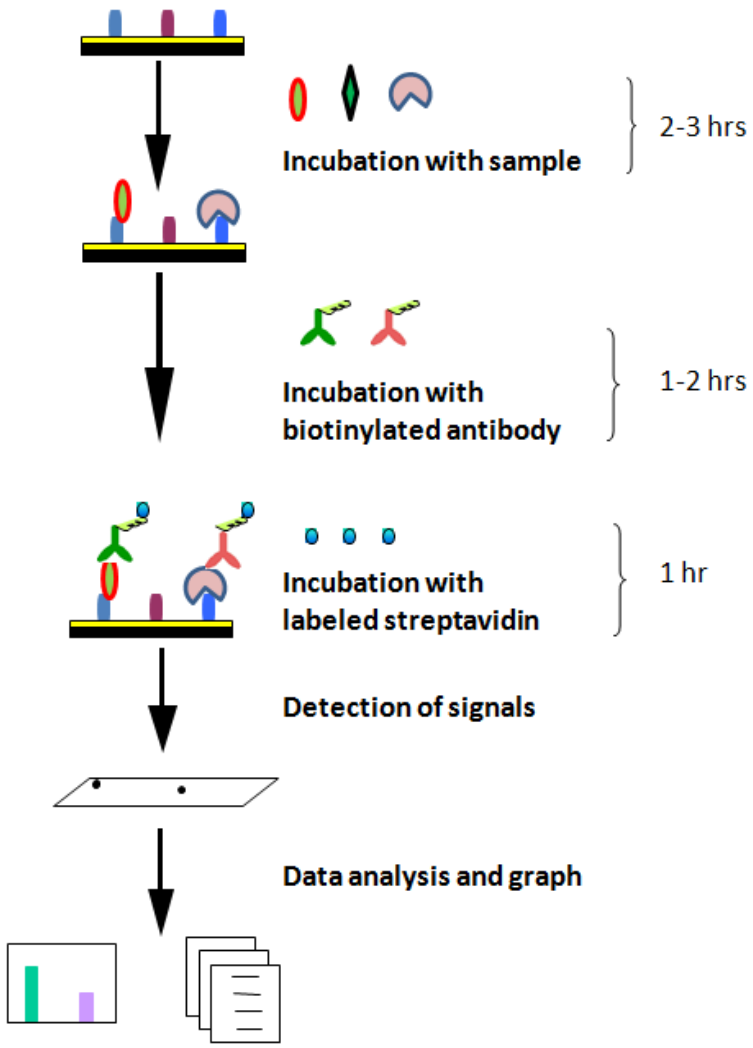
How It Works

Label-based Procedure



Sandwich-based Procedure



II. Materials Provided

Upon receipt, all components of the RayBiotech Glycan Array 100 kit should be stored at -20^{0}C. After initial use, remaining reagents should be stored at 4°C to avoid repeated freeze-thaw cycles and may be stored for up to 3 months (Labeling Reagent, Item B, should be prepared fresh each time before use). Unused glass slides should be kept at -20 °C and repeated freeze-thaw cycles should be avoided (slides may be stored for up to 6 months). The entire kit should be used within 6 months of purchase.

Components

Item	Description	1-Slide kit	2-Slide kit
A	Dialysis Vials	8	16
B	Labeling Reagent	2	4
C	Stop Solution	1	2
D	Glycan Array Glass Slide Assembly*	1 Slide	2 Slides
E	Sample Diluent	1	1
F	20X Wash Buffer I	1	1
G	20X Wash Buffer II	1	1
H	Cy3 equivalent dye-conjugated Streptavidin**	1	2
I	Slide Washer/Dryer	1	1
J	Adhesive device sealer	5	10
K	Labeling Buffer	1	1
L	Floating Dialysis Rack	1	1
M	Manual	1	1

***** Each slide contains 4 identical subarrays
****** HiLyte Plus™ 555

Additional Materials Required

- Detection antibodies of interest (For sandwich-based method only)
- Distilled or de-ionized water
- Orbital shaker
- Laser scanner for fluorescence detection

- Aluminum foil
- Small plastic or glass containers
- 1.5 mL Polypropylene microcentrifuge tubes
- KCl, NaCl, KH_2PO_4 and Na_2HPO_4 (For label-based method only)
- Beaker, stir plate and stir bar
- Pipettors, pipette tips and other common lab consumables

III. General Considerations

A. Label-Based vs. Sandwich-Based Method

The RayBiotech Glycan Array 100 Kit can be used with either a label-based method or with a sandwich-based method.

- In the label-based method, the proteins or antibodies in the sample are biotin labeled (via a simple reaction targeting primary amines), allowing direct detection on the array via a cy3 equivalent dye-conjugated biotin-streptavidin complex. A complete protocol and the primary materials for this procedure are included with the kit.
- The sandwich-based method is used for antibody-based detection of target proteins captured on the array. The user will need to supply the labeled reporter antibodies specific for their protein of interest. An example protocol for this procedure with a general "Antibody Cocktail" is included in this manual. Specific antibody concentrations and conditions will need to be determined by the end user.

B. Preparation of Samples

- We recommend the following parameters for your samples:
    - 300 to 400 µl of 40X diluted serum, plasma, cell culture media, or other body fluid, or 50-500 µg/ml of protein for cell and tissue lysates.

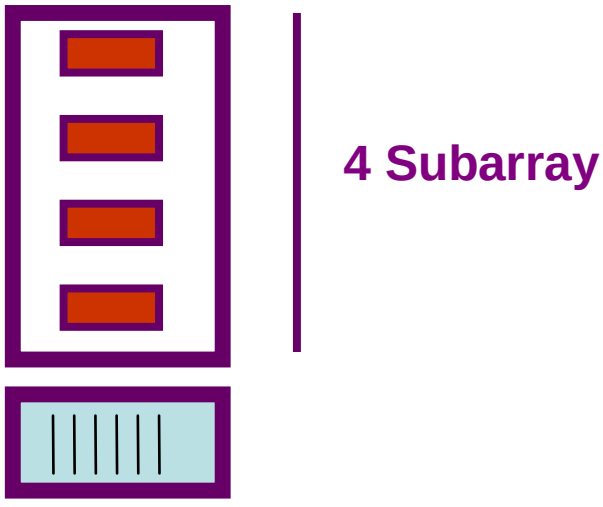
Note: If you experience high background or the readings exceed the detection range, further dilution of your sample is recommended.

C. Handling Glass Slides

- The microarray slides are delicate. Please do not touch the surface of the slides with pipette tips, forceps or your fingers. Hold the slides by the edges only.
- Handle the slides with latex free gloves in a clean environment.
- Do not remove the glass slide from the chamber assembly until step 21, and take great care not to break the glass slide when doing so.
- Because there is no barcode on the slide, transcribe the slide serial number from the slide bag to the back of the slide with a permanent marker before discarding the slide bag. Once the slide's frame is removed, this will allow you to distinguish one slide from another.
- Remove reagents/sample by gently applying suction with a pipette to corners of each chamber. Do not touch the printed area of the array, only the sides.

D. Layout of Glycan Array 100 Glass Slide

Two or four identical sub-arrays on one slide



E. Incubations and Washes

- Cover the incubation chamber with adhesive film during incubation to prevent evaporation, particularly when incubation is more than 2 hours.
- Avoid foaming during incubation steps and wash steps. Be sure to remove all bubbles from the sub-array surface.
- Perform all incubation and wash steps with gentle rocking motion (~0.5 to 1 cycle/sec).
- Avoid cross-contamination of samples to neighboring wells. To remove Wash Buffers and other reagents from chamber wells, you may invert the Glass Slide Assembly to decant, and aspirate the remaining liquid.
- Several incubation steps such as step 11 (sample incubation), step 15 (detection antibody incubation), may be done overnight at 4^0C. Please make sure to cover the incubation chamber tightly to prevent evaporation.
- Unlike most Cy3 fluors, the HiLyte Plus™ 555 used in this kit is very stable at room temperature (RT) and resistant to photobleaching on the hybridized glass slides. However, please protect glass slides from directly strong light and temperatures above RT.

IV. Protocol

READ ENTIRE PROTOCOL BEFORE STARTING

Note: Biotin Label-Based protocol starts here. For the Sandwich-Based protocol (using researcher's own detection antibody), start at section ***C. Dry the Glass Slide,*** *step 8, on page 11. Do not do steps 1-7.*

A. Dialysis of Sample

Note: Samples must be dialyzed prior to biotin-labeling (Steps 5–7).

1. To prepare dialysis buffer (1X PBS, pH=8.0), dissolve 0.6 g KCl, 24 g NaCl, 0.6 g KH_2PO_4 and 3.45 g Na_2HPO_4 in 2500 ml ddH_2O. Adjust pH=8.0 with 1M NaOH and adjust final volume to 3000 ml with ddH_2O.
2. Add each sample into a separate Dialysis Tube (Item A). Load 200 µl cell culture supernatant or 100 µl cell lysates or tissue lysate (1~2 mg/ml total protein) or 20 µl serum or plasma + 80 µl 1X PBS, pH=8 (5-fold dilution. Carefully place Dialysis Tubes into Floating Dialysis Rack (Item L).

Note: If the samples appear to be cloudy, transfer the samples to a clean tube, centrifuge at 13,000 rpm for 20 minutes at 2-8°C. If the samples are still not clear, store them at -20°C for 20 minutes. Remove from the freezer, immediately centrifuge at 13,000 rpm for 20 minutes at 2-8°C.

3. Place Floating Dialysis Rack into ≥500 ml dialysis buffer in a large beaker. Place beaker on a stir plate and dialyze, for at least 3 hours at 4°C, stirring buffer gently. Then exchange the 1X PBS buffer and repeat dialysis for at least 3 hours at 4°C. Transfer dialyzed sample to a clean eppendorf tube. Spin dialyzed samples for 5 min at 10,000 rpm to remove any particulates or precipitates, and then transfer the supernatants to a clean tube.

Note: The sample volume may change during dialysis.

Note: Dialysis procedure may proceed overnight.

Note: Determine the total protein concentration for cell culture supernatants or cell/tissue lysate after dialysis procedure (Step 3). We recommended using a BCA total protein assay (eg, RayBiotech, Catalog # 68QT-BCAPro-S1000).

B. Biotin-labeling Sample

Note: Amines (e.g., Tris, glycine) and azides quench the biotinylation reaction. Avoid contaminating samples with these chemicals prior to biotinylation.

4. Immediately before use, prepare 1X Labeling Reagent. Briefly spin down the Labeling Reagent tube (Item B). Add 100 µl 1X PBS into the tube, pipette up and down or vortex slightly to dissolve the lyophilized reagent.

5. Add 1X Labeling Reagent to dialyzed samples.

    a. For labeling cell culture supernatants: transfer 180 µl dialyzed sample into a new tube. Add 36 µl of 1X Labeling Reagent Solution per 1 mg total protein in dialyzed cell culture supernatant. Mix well. For example, if sample's total protein concentration is 0.5 mg/ml you need to add 3.24 µl 1X Labeling Reagent to the tube of 180 µl dialyzed sample.

Note: You need to biotin-label 360 µl of dialyzed sample if dilution of the biotin-labeled samples is 2 fold in step 11 on page 12.

    b. For labeling serum or plasma: Add 22 µl of 1X Labeling Reagent Solution into a new tube containing 35 µl dialyzed serum or plasma sample and 155 µl Labeling Buffer (Item K).

    c. For labeling cell or tissue lysates: transfer 30 µg (15 µl of 2 mg/ml) cell or tissue lysates into a tube and add Labeling Buffer (Item K) for a total volume of 300 µl. Then add 3.3 µl of 1X Labeling Reagent Solution.

Note: To normalize serum/plasma or cell/tissue lysate concentrations during biotinylation, measure sample volume before and after dialysis. Then

adjust the volumes of dialyzed serum/plasma or cell/tissue lysates and Labeling Buffer to compensate. For example, if the sample volume doubles after dialysis, then use twice as much serum/plasma in the labeling reaction (70 µl) and reduce the Labeling Buffer to 120 µl.

6. Incubate the reaction solution at room temperature with gentle rocking or shaking for 30 min. Mix the reaction solution by gently tapping the tube every 5 min.
7. Add 3 µl Stop Solution (Item C) into each reaction tube and immediately dialyze as directed in Steps 1–3 on pages 9-10.

Note: Biotinylated samples can be stored at -20°C or -80°C until you are ready to proceed with the assay.

C. Dry the Glass Slide

Note: Sandwich-Based protocol starts here.

8. Take out the package containing the Glycan Array Glass Slide Assembly (Item D) and let the slide equilibrate to room temperature inside the sealed plastic bag for 20-30 minutes. Remove slide from the plastic bag; peel off the cover film, and let it air dry at room temperature for another 1-2 hours. Do not disassemble the Glass Slide from the chamber assembly.

Note: Protect the slide from dust or other contaminants.

D. Blocking and Incubation

Note: Glass slide should be <u>completely</u> dry before adding Sample Diluent to wells.

9. Block sub-arrays by adding 400µl Sample Diluent (Item E) into each well and incubate at room temperature for 30 min. Ensure there are no bubbles on the array surface.

10. Immediately prior to sample incubation, spin biotin-labeled samples for 5 min at 10,000 rpm to remove any particulates or precipitates. Dilute samples with Sample Diluent (Item E).

Note: Recommended dilution of the biotin-labeled samples with Sample Diluent prior to incubation is 2-10 fold for cell culture supernatants, 20-fold for serum/plasma or 30 fold cell/tissue lysate.

11. Decant buffer from each well. Add 400µl of sample to each well. Incubate arrays with gentle rocking or shaking at room temperature for 2-3 hours. *(Longer incubation time is preferable if higher signal intensity is desired)*

Note: This step may be done overnight at 4°C for highest intensities.

12. Wash:

    a. Based on number of samples and remaining protocol, calculate the amounts of 1x Wash Buffers I & II that are needed for each step of the protocol. Separately dilute required amounts of 20x Wash Buffer I and 20x Wash Buffer II with ddH_2O to 1x concentration. *For example if 12 ml of 1x Wash Buffer I is needed then 600 µl of 20x Wash Buffer I would be diluted to a final volume of 12 ml.*

    b. Decant the samples from each well, and wash each well 5 times (5 min each) with 800 µl of 1x Wash Buffer I at room temperature with gentle shaking. Completely remove wash buffer between each wash step.

    c. (*Optional for Cell and Tissue Lysates*) Put the glass slide with frame into a box with 1x Wash Buffer I (cover the whole glass slide and frame with Wash Buffer I), and wash at room temperature with gentle shaking for 20 min.

    d. Decant the 1x Wash Buffer I from each well, wash 2 times (5 min each) with 800 µl of 1x Wash Buffer II at room temperature with

gentle shaking. Completely remove wash buffer between each wash step.

Note: Incomplete removal of the wash buffer after each wash step may cause "dark spots". (i.e., background signal higher than that of the spot.)

E. Incubation with Biotinylated Detection Antibody (provided by researcher)

*Note: For the Label-Based protocol, go directly to **F. Incubation with Cy3 Equivalent Dye-Streptavidin**, step 17, on page 13. Do not do steps 13-16.*

13. If the researcher wishes to use their own antibody to detect specific bound proteins, we recommend using a biotinylated antibody at a dilution appropriate for Western blot. Optimal dilution must be determined by the researcher.

14. Dilute the detection antibody in Sample Diluent. Mix well and spin briefly.

15. Add 400 µl of the detection antibody to each well. Incubate at room temperature for 1-2 hours.

Note: Longer incubation time is preferable for higher signals.

16. Decant the samples from each well, and wash 5 times with 800 µl of 1x Wash Buffer I and then 2 times with 800 µl of 1x Wash Buffer II at room temperature with gentle shaking. Completely remove wash buffer between each wash step.

F. Incubation with Cy3 Equivalent Dye-Streptavidin

17. Prepare 1X Dye-conjugated Streptavidin:
    a) Briefly spin down the Cy3 equivalent dye-conjugated streptavidin tube (Item H) immediately before use.
    b) Add 1000 µl of Sample Diluent to Cy3 equivalent dye-conjugated streptavidin tube. Mix gently (do not store the stock solution for later use).

c) Add 400 µl of Cy3-Conjugated Streptavidin stock solution into a tube with 1400 µl of Blocking Buffer. Mix gently to prepare 1X Cy3-Conjugated Streptavidin

18. Add 400 µl of 1X Cy3 equivalent dye-conjugated streptavidin to each well. Cover the incubation chamber with the plastic adhesive strips (Item J) and cover the slide with aluminum foil to avoid exposure to light or incubate in dark room.

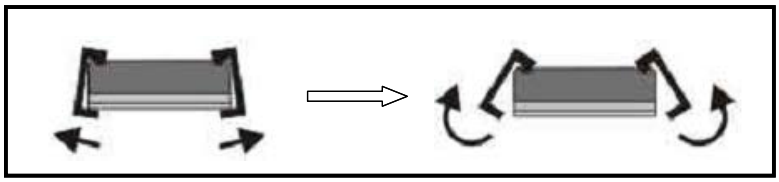
19. Incubate the slide with Cy3-Conjugated Streptavidin at RT for 1 hour with gentle rocking or shaking.

20. Decant the samples from each well, and wash 5 times with 800 µl of 1x Wash Buffer I at room temperature with gentle shaking. Completely remove wash buffer in each wash step.

G. Fluorescence Detection

21. Disassemble the slide assembly by pushing clips outward from the slide side, as shown below. Carefully remove the slide from the gasket.

Note: Be careful not to touch the surface of the array.



22. Gently place the slide in the slide Washer/Dryer (a 4-slide holder/centrifuge tube, Item I), add enough 1x Wash Buffer I (about 30 ml) to cover the whole slide, and then gently shake at room temperature for 15 minutes. Decant Wash Buffer I. Wash with 1x Wash Buffer II (about 30 ml) and gently shake at room temperature for 5 minutes.

23. Finally, wash the glass slide with 30 mL of de-ionized or distilled water for 5 min.

24. Remove water droplets completely by gently applying suction with a pipette. Do not touch the sub-array areas, only the sides of the slide.

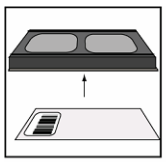
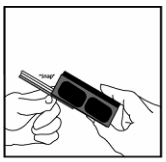
    Make sure the finished glass slide is completely dry before scanning or storage.

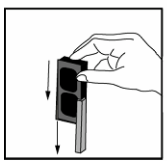
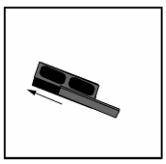
25. Imaging: The signals can be visualized through use of a laser scanner equipped with a Cy3 wavelength such as Axon GenePix. Make sure that the signal from the spot containing the highest concentration receives the highest possible reading, yet remains unsaturated.

Note: ***Unlike most Cy3 fluors, the HiLyte Plus™ Fluor 555 used in this kit is very stable at RT and resistant to photobleaching on completed glass slides. However, please protect glass slides from temperatures above RT and store them in the dark. Do not expose glass slide to strong light, such as sunlight or UV lamp.***

Note: ***If you need to repeat any of the incubation after finishing the experiment, you must first re-assemble the glass slide into the incubation chamber by following step as shown in the figures below. To avoid breaking the printed glass slide, you may first want to practice assembling the device with a blank glass slide.***

1. *Apply slide to incubation chamber barcode facing upward as in image A (below).*
2. *Gently snap one edge of a snap-on side as shown in image B.*
3. *Gently press other of side against lab bench and push in lengthwise direction (image C).*
4. *Repeat with the other side (image D)*

A   B

C   D

H. Data Analysis

Data extraction can be done with most of the microarray analysis software (GenePix, ScanArray Express, ArrayVision, or MicroVigene).
NOTE: Due to the difficulty of printing glycans, each glycan is printed in quadruplicate. We guarantee each glycan to have at least 3 spots present, and additionally guarantee no more than 5 total spots per array will be missing.

V. Glycan Array Map

Glycan structures and molecular weights are available at this link:
www.raybiotech.com/GlycanArray100Structures

	100 Glycan Array Map																											
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27
1	POS1	POS2	POS3	NEG	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23
2	POS1	POS2	POS3	NEG	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23
3	POS1	POS2	POS3	NEG	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23
4	POS1	POS2	POS3	NEG	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23
5	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50
6	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50
7	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50
8	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	50
9	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77
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11	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77
12	51	52	53	54	55	56	57	58	59	60	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77
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14	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100	NEG	POS3	POS2	POS1
15	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100	NEG	POS3	POS2	POS1
16	78	79	80	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97	98	99	100	NEG	POS3	POS2	POS1

1	β-Glc-	51	GlcNAc-β-1,4-GlcNAc-β-
2	β-Gal-	52	β-D-GlcA-
3	α-Man-	53	Gal-β-1,4-(6S)GlNAc-β-
4	α-Fuc-	54	GlcNAc-α-1,3-(Glc-α-1,2-Glc-α-1,2)-Gal-α-1,3-Glc-α-
5	α-Rha-	55	Gal-β-1,3-GalNAc-β-1,4-(Neu5Gc-α-2,3)-Gal-β-1,4-Glc-β-
6	β-GlcNAc-	56	Sisomicin Sulfate
7	β-GalNAc-	57	GalNAc-α-1,3-(Fuc-α-1,2)-Gal-β- [Blood A antigen trisaccharide]
8	Tobramycin	58	Fuc-α-1,2-Gal-β-1,4-GlcNAc-β- [Blood H antigen trisaccharide]
9	Gal-β-1,3-GlcNAc-β-	59	Gal-α-1,3-(Fuc-α-1,2)-Gal-β- [Blood B antigen trisaccharide]

10	Gal-α-1,3-Gal-β-1,3-GlcNAc-β-	60	Fuc-α-1,2-Gal-β-1,3-GlcNAc-β-1,3-Gal-β-1,4-Glc-β- [LNFP I]
11	Neu5Ac-α-2,3-Gal-β-1,3-GlcNAc-β-	61	Fuc-α-1,2-Gal-β-1,4-Glc-β- [Blood H antigen trisaccharide]
12	Neu5Ac-α-2,6-Gal-β-1,3-GlcNAc-β-	62	Gal-α-1,3-(Fuc-α-1,2)-Gal-β-1,4-Glc-β- [Blood B antigen tetrasaccharide]
13	Neu5Gc-α-2,3-Gal-β-1,3-GlcNAc-β-	63	(Fuc-α-1,2)-Gal-β-1,4-(Fuc-α-1,3)-GlcNAc-β- [Lewis Y]
14	Neu5Gc-α-2,6-Gal-β-1,3-GlcNAc-β-	64	(Fuc-α-1,2)-Gal-β-1,3-(Fuc-α-1,4)-GlcNAc-β- [Lewis B]
15	Gal-β-1,3-(Fuc-α-1,4)-GlcNAc-β- [Lewis A]	65	Gal-β-1,3-(Fuc-α-1,4)-GlcNAc-β-1,3-Gal-β-1,4-(Fuc-α-1,4)-Glc-β- [Lewis A]
16	Gal-β-1,4-Glc-β-	66	Gal-β-1,3-GalNAc-β-
17	Gal-α-1,3-Gal-β-1,4-Glc-β-	67	Gal-β-1,3-(Neu5Ac-α-2,6)-GalNAc-β-
18	Gal-α-1,4-Gal-β-1,4-Glc-β-	68	Neu5Ac-α-2,6-Gal-β-1,3-GalNAc-β-
19	GlcNAc-β-1,3-Gal-β-1,4-Glc-β-	69	Neu5Ac-α-2,6-Gal-β-1,3-(Neu5Ac-α-2,6)-GalNAc-β-
20	GalNAc-β-1,3-Gal-β-1,4-Glc-β-	70	Neu5Ac-α-2,3-Gal-β-1,3-(Neu5Ac-α-2,6)-GalNAc-β-
21	Neu5Ac-α-2,3-Gal-β-1,4-Glc-β-	71	Neu5Ac-α-2,6-(Neu5Ac-α-2,3)-Gal-β-1,3-GalNAc-β-
22	Neu5Ac-α-2,6-Gal-β-1,4-Glc-β-	72	GalNAc-β-1,4-(Neu5Ac-α-2,3)-Gal-β-1,4-Glc-β- [GM2]
23	Neu5Gc-α-2,3-Gal-β-1,4-Glc-β-	73	GalNAc-β-1,4-(Neu5Ac-α-2,8-Neu5Ac-α-2,3)-Gal-β-1,4-Glc-β- [GD2]
24	Neu5Ac-α-2,6-Gal-β-1,4-Glc-β-	74	Gal-α-1,4-Gal-β-1,4-GlcNAc-β-
25	Gal-β-1,4-(Fuc-α-1,3)-Glc-β-	75	β-D-Rha-
26	GalNAc-β-1,3-Gal-α-1,4-Gal-β-1,4-Glc-β-	76	Glc-α-1,4-Glc-β-
27	GlcNAc-β-1,6-GlcNAc-β-	77	Glc-α-1,6-Glc-α-1,4-Glc-β-
28	4-P-GlcNAc-β-1,6-GlcNAc-β-	78	Maltotriose-β-
29	Glc-α-1,2-Gal-α-1,3-Glc-α-	79	Glc-α-1,6-Glc-α-1,6-Glc-β-
30	Gal-β-1,3-GalNAc-α-	80	Maltotetraose-β-
31	Gal-β-1,4-GlcNAc-β-	81	GlcNAc-α-1,4-GlcA-β-1,4-GlcNAc-α1,4-GlcA-β-
32	Gal-β-1,4 -(Fuc-α-1,3)-GlcNAc-β- [Lewis X]	82	Maltohexaose-β-
33	Neu5Ac-α-2,3-Gal-β-1,4-(Fuc-α-1,3)-GlcNAc-β- [Sialyl Lewis X]	83	Maltoheptaose-β-
34	Neu5Ac-α-2,3-Gal-β-1,3 -(Fuc-α-1,4)-GlcNAc-β- [Sialyl Lewis A]	84	Acarbose-β-
35	Neu5Gc-α-2,3-Gal-β-1,3-(Fuc-α-1,4)-GlcNAc-β- [Sialyl Lewis A]	85	D-pentamannuronic acid-β-
36	Gal-α-1,4-Gal-β-1,3-GlcNAc-β-	86	L-pentaguluronic acid-β-

37	Gal-β-1,4-GlcNAc-β-1,3-Gal-β-1,4-Glc-β- [LNnT]	87	D-cellose-β-
38	GlcA-β-1,4-GlcNAc-α-1,4-GlcA-β-	88	Gal-α-1,3-Gal-β-
39	GlcNAc-β-1,6-(Gal-β-1,3)-GalNAc-α-O-Ser	89	β-1,4-Xylotetrose
40	Neu5Ac-α-2,3Gal-β-1,4-(6S)GlcNAc-β-	90	Chitin-trisaccharide
41	GalNAc-β-1,4-GlcNAc-β-	91	KDN-α-2,8-Neu5Ac-α-2,3-Gal-β-1,4-Glc-β-
42	Neu5Ac-α-2,8-Neu5Ac-α-2,3-Gal β-1,4-Glc-β-	92	Neu5Ac-α-2,8-Neu5Gc-α-2,3-Gal-β-1,4-Glc-β-
43	Neu5Gc-α-2,8-Neu5Ac-α-2,3-Gal-β-1,4-Glc-β-	93	Neu5Ac-α-2,8-Neu5Ac-α-2,8-Neu5Ac-α-2,3-Gal-β-1,4-Glc-β-
44	GalNAc-α-1,3-(Fuc-α-1,2)-Gal-β-1,4-Glc-β- [Blood A antigen tetrose]	94	Neu5Ac-α-2,8-Neu5Ac-α-2,8-Neu5Ac-α-2,8 Neu5Ac-α-2,3-Gal β-1,4-Glc-β-
45	GlcNAc-β-1,2-Man-α-	95	Gal-β-1,3-GalNAc-β-1,4-(Neu5Ac-α-2,3)-Gal-β-1,4-Glc-β-
46	Neu5Ac-α-2,3-Gal-β-	96	Gentamicin Sulfate
47	Gal-β-1,3 -GalNAc-β-1,3-Gal-β-	97	Kanamycin sulfate
48	Glc-α-1,2-Gal-α-	98	Geneticin Disulfate Salt (G418)
49	Gal-β-1,4-(Fuc-α-1,3)-GlcNAc-β-1,3-Gal-β-	99	Neomycin trisulfate
50	Neu5Ac-α-2,3-Gal-β-1,4-(Fuc-α-1,3)-Glc-β- [3-Sialyl-3-fucosyllactose/ F-SL]	100	SGP

VI. Interpretation of Results and Two Examples:

A. Explanation of Controls Spots

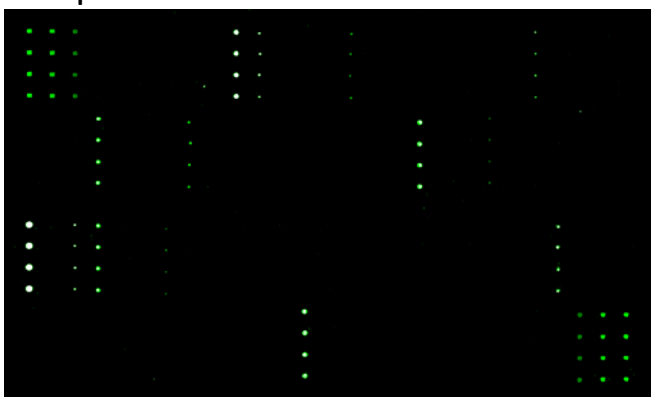
1) **Positive Control spots (POS1, POS2, POS3)** are standardized amounts of biotinylated IgGs printed directly onto the array. All other variables being equal, the Positive Control intensities will be the same for each sub-array. This allows for normalization based upon the relative fluorescence signal responses to a known control, much as "housekeeping" genes or proteins are used to normalize results in PCR or Western blots, respectively.

2) **Negative Control (NEG)** spots contain a protein-containing buffer (used to dilute glycans printed on the array). Their signal intensities represent non-specific binding of Biotin-conjugated anti-Cytokines and/or the Cy3-Conjugated Streptavidin. Negative control signal intensities are usually very close to background signals in each sub-array.

B. Typical Results Obtained with RayBio® Glycan Array 100

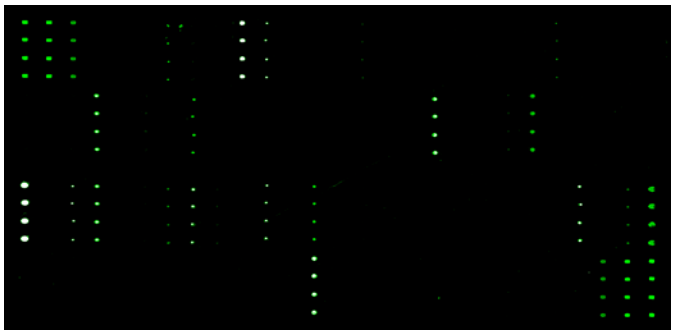
The following figure shows the RayBio® Glycan Array 100 probed with biotin-labeled lectin mixtures. The images were captured using a Axon GenePix laser scanner. The strong signals in the upper left and lower right corners of each array are Positive Controls, which can be used to identify the orientation and help normalize the results between arrays. In sample 1, glycans #6, #7, #27, #51, #54, and #90 showed strong binding activities to biotin-lectins (b-BSL I, b-WGA) indicating that those glycans contain aGal, aGalNAc, and GlcNAc groups in their structures. In sample 2, glycans #3, #50, #58, and #61 showed stronger signals than in sample 1 meaning that these glycans have aFuc groups in their structures.

Sample 1



Biotin-lectin mixture:
b-BSL I, b-WGA (0.4 µg/mL)

Sample-2



Biotin-lectin mixture: b-BSL I, b-WGA, b-UEA I (0.4 µg/mL) and b-Con A: (0.04 µg/mL)

Blank control

Sample Diluent only (Item E)

If scanned using optimal settings, 3 distinct signal intensities will be seen: POS1>POS2>POS3. If all of these signals are of similar intensity, try increasing or decreasing laser power and/or signal gain settings.

C. Background Subtraction:

Once you have obtained fluorescence intensity data, you should subtract the background and normalize to the Positive Control signals before proceeding to analysis.

Most laser fluorescence scanner software has an option to automatically measure the local background around each spot. For best results, we recommend comparing signal intensities representing the MEDIAN background signals minus local background. If your resulting fluorescence signal intensity reports do not include these values (e.g., a column labeled as “MED532-B532”), you may need to subtract the background manually or change the default settings on your scanner’s data report menu.

D. Normalization of Array Data:

To normalize signal intensity data, one sub-array is defined as "reference" to which the other arrays are normalized. This choice is arbitrary. For example, in our Analysis Tool Software (described below), the array represented by data entered in the left-most column each worksheet is the default “reference array.”

You can calculate the normalized values as follows:

X(Ny) = X(y) * P1/P(y)

Where:
P1 = mean signal intensity of POS spots on reference array
P(y) = mean signal intensity of POS spots on Array "y"
X(y) = mean signal intensity for spot "X" on Array "y"
X(Ny) = normalized signal intensity for spot "X" on Array "y"

The RayBio® Analysis Tool software is available for use with data obtained using RayBio® Glycan Array 100. You can copy and paste your signal intensity data (with and without background) into the Analysis Tool, and it will automatically normalize signal intensities to the Positive Controls.

To order the Analysis Tool, please contact us at +1-770-729-2992 or info@raybiotech.com for more information.

E. Threshold of significant difference in samples:

After subtracting background signals and normalization to Positive Controls, comparison of signal intensities between and among array images can be used to determine relative differences between samples or groups.

Any $\geqslant$1.5-fold increase or $\leqslant$0.65-fold decrease in signal intensity for a single analyte between samples or groups may be considered a measurable and significant difference in expression, provided that both sets of signals are well above background (Mean background + 2 standard deviations, accuracy $\approx$ 95%).

VII. Troubleshooting Guide

Problem	Cause	Recommendation
Weak Signal	Inadequate detection	Increase laser power and PMT parameters
	Inadequate reagent volumes or improper dilution	Check pipettes and ensure correct preparation
	Short incubation time	Ensure sufficient incubation time or change sample incubation step to overnight
	Too low glycan concentration in sample	Reduce amount of dilution or concentrate sample
	Improper storage of kit	Store kit as suggested temperature; Don't freeze/thaw the slide
Uneven Signal	Bubble formed during incubation	Handle and pipette solutions more gently; De-gas solutions prior to use
	Arrays are not completed covered by reagent	Prepare more reagent and completely cover arrays with solution
	Reagent evaporation	Cover the incubation chamber with adhesive film during incubation
General	Cross-contamination from neighboring wells	Avoid overflowing wash buffer
	Comet tail formation	Air dry the slide for at least 1 hour before usage
	Inadequate detection	Increase laser power that the highest concentration for each lectin receives the highest possible reading yet remains unsaturated
High Background	Overexposure	Lower the laser power
	Dark spots	Completely remove wash buffer in each wash step
	Insufficient wash	Increase wash time and use more wash buffer
	Dust	Minimize dust in work environment before starting experiment
	Slide is allowed to dry out	Take additional precautions to prevent slides from dying out during experiment

Note:

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